Sesquiterpene–Flavanone [4+2] Hybrids, Syzygioblanes A–C, Found in an Indonesian

Traditional Medicine, "Jampu

Cyclic

Salo" (Syzygium oblanceolatum)

by PDTI Master Turnitin

Submission date: 04-Jul-2024 02:44PM (UTC+0800)

Submission ID: 2412396961
File name: Inter_1.pdf (2.5M)

Word count: 4657

Character count: 23297



pubs.acs.org/OrgLett Letter

Cyclic Sesquiterpene—Flavanone [4+2] Hybrids, Syzygioblanes A—C, Found in an Indonesian Traditional Medicine, "Jampu Salo" (Syzygium oblanceolatum)

Nona Koga, Yohei Saito, Katsunori Miyake, Saidanxia Amuti, Shuichi Fukuyoshi, Satoshi Yoshida, Sota Sato, Yusuke Yamada, Akihito Ikeda, Naruhiko Adachi, Masato Kawasaki, Akira Takasu, Shinji Aramaki, Toshiya Senda, Abdul Rahim, Ahmad Najib, Gemini Alam, Nobuyuki Tanaka, and Kyoko Nakagawa-Goto*



Cite This: Org. Lett. 2024, 26, 4302-4307



ACCESS

Metrics & More

Article Recommendations

s Supporting Information

ABSTRACT: A plant used in an Indonesian traditional herbal medicine as a diabetes treatment and known locally as "Jampu Salo" was collected on Sulawesi Island, Indonesia. It was identified as Syzygium oblanceolatum (C. B. Rob.) Merr. (Myrtaceae) and found for the first time in Sulawesi; it was previously reported only in the eastern Philippines and Borneo. A phytochemical study of S. oblanceolatum led to the isolation of three unprecedented meroterpenoids, syzygioblanes A–C (1–3, respectively). These compounds might be biosynthesized through [4+2] cycloaddition of various germacrane-based cyclic sesquiterpenoids with the flavone desmethoxymatteucinol to form a spiro skeleton. The unique and complex structures were elucidated by microcrystal electron diffraction analysis in addition to general analytical

Indonesian medicinal plant,
"Jampu Salo"
Syzygium oblanceolatum
collected in Sulawesi

Cys

[4+2]
HO
H
H
H
H
H
H
H
H
H
H
H
H
Spiro-ring
formation
Desmethoxymatteucinol (4)

Syzygioblanes A-C (1-3)

techniques such as high-resolution mass spectrometry, various nuclear magnetic resonance methods, and infrared spectroscopy. Synchrotron X-ray diffraction and calculations of electronic circular dichroism spectra helped to determine the absolute configurations. The newly isolated compounds exhibited collateral sensitivity to more strongly inhibit the growth of a multidrug resistant tumor cell line compared to a chemosensitive tumor cell line.

"Jampu Salo" is the local name for a leaf decoction used in Indonesian traditional folk medicine to treat diabetes. The plant was collected on Sulawesi Island in 2018 and identified as Syzygium oblanceolatum by taxonomist Dr. Wuu-Kuang Soh. This report is the first record of S. oblanceolatum from Sulawesi, known previously only in the Eastern Philippines and Borneo.¹

The plant genus Syzygium, distributed in subtropical to tropical regions, contains numerous species that contribute significantly to the diversity of tropical forests, especially in Southeast Asia. Its enormous morphological and ecological variations make phylogenetic elucidation difficult, although ~1200 species have been identified to date. Syzygium aromaticum (L.) Merr. & L. M. Perry, the best-known species, is traditionally used in East Asia as a spice (clove) in cooking, as a treatment for burns and wounds, and as an analgesic in dentistry. Syzygium jambos (L.) Alston and Syzygium fibrosum (F. M. Bailey) T. G. Hartley & L. M. Perry are widely used as foods, such as jams and jellies, while Syzygium cordatum Hochst ex Krauss, Syzygium samarangense (Blume) Merr. & L. M. Perry, S. aromaticum, and Syzygium cumini (L.) Skeels are

used to treat cancer, fever, diarrhea, and diabetes. Although the genus *Syzygium* has many medicinal properties and is used in traditional remedies, surprisingly few species have been evaluated phytochemically, and there are no phytochemical reports regarding *S. oblanceolatum*.

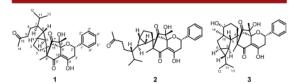


Figure 1. Structures of syzygioblanes A-C (1-3, respectively).

Received: April 8, 2024 Revised: May 3, 2024 Accepted: May 8, 2024 Published: May 10, 2024





Our phytochemical study of *S. oblanceolatum* resulted in the identification of three structurally and biologically unique meroterpenoids, syzygioblanes A–C (1–3, respectively),³ which are hybrids of a germacrane-based cyclic sesquiterpene and a flavanone (Figure 1).

Compound 1 was obtained as a colorless amorphous solid with a molecular formula of $C_{32}H_{40}O_6$ established from a protonated molecule peak at m/z 521.2893 [M + H]⁺ (calcd 521.2903) by HRMS. IR absorptions at 1717, 1663, and 1603 cm⁻¹ implied the presence of carbonyl(s) and an α,β -unsaturated ketone. The ¹H NMR spectrum (Table 1)

Table 1. ¹H NMR Data for Compounds 1-3 (600 MHz, CDCl₃)

	$\delta_{ m H}$				
position	1	2	3		
1	0.97 m	1.18 m	3.16 dd (11.2, 4.4)		
2	1.65 m	1.70 m	1.49 m, 1.56 m		
3	0.94 m, 1.19 m	1.24 m, 1.72 m	1.15 m, 1.56 m		
5	0.71 dd (6.3, 3.4)	0.97 dd (6.0, 3.4)	0.65 d (5.7)		
6	0.56 ddd (10.0, 3.4, 3.2)	0.34 ddd (10.1, 3.4, 3.4)	0.56 m		
7	0.41 m	0.50 m	0.49 m		
8	1.41 m, 1.62 m	1.56 m, 1.65 m	1.10 m		
9	2.37 m, 2.52 m	2.40 m, 2.52 m	0.75 m, 1.72 m		
11	1.82 m	1.66 m	1.25 m		
12	0.88 d (6.9)	0.93 d (6.9)	1.02 d (6.8)		
13	0.91 d (6.9)	0.88 d (6.9)	0.83 d (6.8)		
14	2.13 s	2.14 s	0.74 s		
15	1.40 d (12.8), 2.55 d (12.8)	2.22 d (12.8), 2.67 d (12.8)	1.73 d (12.2), 2.28 d (12.2)		
2'	5.20 dd (8.4, 4.1)	5.13 dd (8.6, 4.8)	5.29 dd (6.9, 4.4)		
3'	2.69 m	2.61 m	2.77 m, 2.85 m		
2", 6"	7.39 m	7.39 m	7.44 m		
3", 5"	7.40 m	7.38 m	7.39 m		
4"	7.34 m	7.34 m	7.35 m		
4'-OH	12.6 brs	12.6 brs	12.9 brs		
6'-Me	1.24 s	1.16 s	1.16 s		
8'-Me	1.43 s	1.44 s	1.40 s		
8'-OH			2.45 brs		

indicated the presence of a deshielded oxymethine [$\delta_{\rm H}$ 5.20 (1H, dd)], five methyls [$\delta_{\rm H}$ 0.91, 0.88 (each 3H, d), $\delta_{\rm H}$ 1.43, 1.24 (each 3H, s)], including a deshielded methyl [$\delta_{\rm H}$ 2.13 (3H, s)], a monosubstituted benzene [$\delta_{\rm H}$ 7.40, 7.39 (each 2H, m), 7.34 (1H, m)], and a hydrogen-bonded hydroxy group ($\delta_{\rm H}$ 12.6). The ¹³C NMR spectrum suggested 32 carbons (Table 2); additional observation combined with DEPT135 and HMQC spectra assigned them as three ketone carbonyl carbons (δc 209.4, 208.6, 196.0), eight sp² carbons, including one oxygenated (δc 168.8) and six aromatic [δc 140.1, 128.7 (overlapped), 128.5, 126.5 (overlapped)], four quaternary carbons (δ c 66.2, 49.8), including two oxygenated (δ c 78.2, 76.4), six methines (δ c 47.3, 32.0, 30.6, 24.8, 22.7), including an oxymethine (δc 73.8), six methylenes (δc 45.2, 42.6, 35.0, 31.5, 24.6, 23.6), and five methyls (δc 29.9, 22.3, 21.0, 17.9, 6.9). A ¹H-¹H COSY correlation of H-2'/H-3', HMBC correlations from H-2' to C-9'/C-1"/C-2", H-3' to C-4'/C-10', and the protons of CH₃-6' to C-5'/C-6'/C-7' and CH₃-8' to C-7'/C-8'/C-9', and H2BC correlations between H-3"/C-2" and H-4"/C-3" implied that compound 1 has a 5,7-dioxo-6,8-dimethylflavanone skeleton (Figure 2).

Table 2. ¹³C NMR Data for Compounds 1-3 (150 MHz, CDCl₃)

		$\delta_{ m C}$	
position	1	2	3
1	22.7	28.0	77.6
2	24.6	28.8	27.8
3	31.5	35.7	33.4
4	66.2	68.5	68.8
5	32.0	34.4	56.5
6	24.8	22.8	24.3
7	47.3	46.9	48.8
8	23.6	25.5	23.0
9	42.6	42.1	44.2
10	209.4	209.2	60.0
11	30.6	31.3	30.3
12	21.0	19.5	21.6
13	17.9	19.3	20.9
14	29.9	30.0	16.4
15	45.2	44.5	37.9
2'	73.8	74.4	73.3
3'	35.0	36.3	34.0
4'	168.8	169.3	169.2
5'	196.0	196.2	194.3
6'	49.8	50.5	43.3
7'	208.6	208.4	207.6
8'	78.2	76.4	76.4
9'	76.4	77.8	77.6
10'	108.3	108.4	110.1
1"	140.1	140.2	140.2
2", 6"	126.5	126.1	126.9
3", 5"	128.7	128.8	128.7
4"	128.5	128.5	128.6
6'-Me	6.9	7.4	6.8
8'-Me	22.3	22.6	22.3



Figure 2. Key correlations of COSY, HMBC, and H2BC for 1.

In addition, the presence of a cyclopentane was suggested by the HMBC cross peaks between H-2/C-1 and C-5 and H-5/C-3, as well as the ¹H-¹H COSY correlation of H-2/H-3. Furthermore, the H2BC correlation of H-5/C-1 in addition to the ¹H-¹H COSY correlations of H-1/H-6/H-5 indicated the presence of a cyclopropane, which was fused to the cyclopentane to form a bicyclic system, a bicyclo[3.1.0]hexane. Moreover, the HMBC correlations from H-15 to C-9'/C-10'/ C-6'/C-3/C-4/C-5, H-3 to C-6', and CH₃-6' to C-4 indicated an unusual carbon-carbon framework containing a spiro[4.5]decane (C-4 is the spirocarbon) and a bicyclo[2.2.2]octane (C-4 and C-15 are bridge carbons). The ¹H-¹H COSY correlations of H-6/H-7, H-8/H-9, and H-11/H-12 and H-13 and the HMBC correlations from H-13 to C-7, H-11 to C-8, and H-14 to C-10 and C-9 supported the presence of isopropyl and butanone fragments at C-7, which, in turn, was connected to C-6. These data indicated that compound 1 is a

meroterpenoid, specifically a 6,8-dimethylflavonoid hybridized with a sesquiterpenoid. A hydroxy group at C-8' was determined from an OH/C-7' HMBC correlation measured in DMSO-d₆. This extremely unique skeleton was further verified by microcrystal electron diffraction (MicroED) analysis (Figure 3). The relative configuration of 1 was

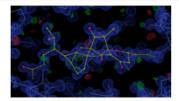


Figure 3. Structure of syzygioblane A (1) from MicroED CryoEM.

deduced from the combined NOESY correlations of H-1/H-5/ H-7/H-9a/H-9b, H-6/CH₃-6', and H-2'/CH₃-8' (Figure 4),

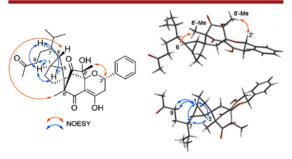


Figure 4. Key NOESY correlations for syzygioblane A (1).

as well as MicroED analysis. Additionally, synchrotron X-ray diffraction analysis with a Flack parameter of 0.07(5) (Figure 5) and ECD calculations (Figure 6) established the absolute configuration of 1 (syzygioblane A).

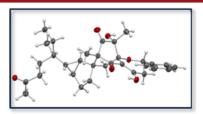


Figure 5. ORTEP view of syzygioblane A (1) showing a 50% probability thermal motion ellipsoid.

Compound 2 was isolated as a colorless, amorphous solid. Compounds 1 and 2 have the same molecular formula $(C_{32}H_{40}O_6)$ based on a protonated molecule peak at m/z 521.2871 [M + H]⁺ (calcd 521.2903) in the HRFABMS of 2. Similarly, on the basis of the one-dimensional (Tables 1 and 2) and two-dimensional NMR (Figure 7) spectra, like 1, compound 2 was also expected to be a meroterpenoid with the same flavanone and bicyclic sesquiterpene components. However, the NOESY correlations of H-1/H-9, H-1/H-7, H-7/H-5, CH₃-8'/H-2', and H-6/H-15 suggested a different stereochemistry at C-4 (Figure 8). An ECD calculation (Figure

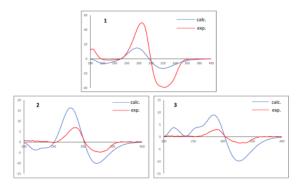


Figure 6. Measured and calculated ECD for 1-3.

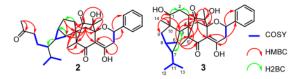


Figure 7. Key COSY, HMBC, and H2BC correlations for 2 and 3.

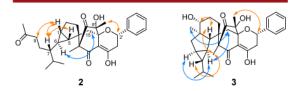


Figure 8. Key NOESY correlations for 2 and 3.

6) and synchrotron X-ray diffraction analysis indicated the absolute configuration of 2 with a Flack parameter of 0.12(9) (Figure S25). All of the data presented above indicated that compound 2 (syzygioblane B) is the C-4 epimer of 1.

Compound 3 (syzygioblane C) was isolated as a pale yellow oil and showed a protonated molecule peak at m/z 521.2871 [M + H]⁺ (calcd 521.2903) corresponding to the molecular formula C32H40O6. The 1H and 13C NMR (Tables 1 and 2) spectra of 3 displayed signal patterns similar to those of 1 and 2 for the flavanone skeleton, the C-2' to C-5', C-7' to C-10', and C-1" to C-6" positions, as well as 4'-OH and 6',8'-CH3. The ¹H-¹H COSY and HMBC correlations (Figure 7) also supported the presence of the same 6,8-dimethylflavanone moiety found in 1 and 2. Signals for an isopropyl unit, C-11 to C-13, were also observed in the NMR spectra of 3. The NMR signals for other portions of 3 were dissimilar from those of 1 and 2, and an additional broad proton was seen at 2.45 ppm, suggesting that compound 3 is also a meroterpenoid containing a 6,8-dimethylflavanone but hybridized with a type of sesquiterpene different from those in 1 and 2. The ¹H-¹H COSY for the sesquiterpene side showed correlations of H-8/H-6, H-7, and H-9, of H-7/H-11, and of H-11/H-12 and H-13 (Figure 7). Additional HMBC correlations were found from H-14 to C-1 (shifted downfield at δ_C 77.6), C-5, C-10, and C-9, H-5 to C-4, and H-15 to C-3 and C-4, as well as H2BC correlations between H-1/C-2, H-3/C-2, and H-6/ C-5 and C-7. Together, all of the data suggested a tricyclic sesquiterpene with a 6-5-3 ring system, a hydroxy moiety at C-1, an isopropyl unit at C-7, and a methyl group at C-10. The

Table 3. Antiproliferative Activity Data of Syzygioblanes A-C (1-3, respectively) versus Different Cell Lines^a

	$IC_{30} (\mu M)^b$					SIc
	A549	MDA-MB-231	MCF-7	KB	KB-VIN	KB/KB-VIN
1	7.7	12.0	10.4	5.8	0.4	14.5
2	7.0	13.0	4.4	5.8	0.6	9.7
3	12.1	33.6	26.6	18.1	4.1	4.4
4	>40	>40	>40	>40	>40	-
PXL^{d} (nM)	3.2	15.0	13.2	3.0	2372.2	0.001

"Abbreviations: A549, lung adenocarcinoma; MDA-MB-231, triple-negative breast cancer (ER-, PR-, and HER2-negative); MCF-7, HER2-negative breast cancer; KB, HeLa derivative, originally isolated from epidermoid; KB-VIN, vincristine resistant KB subline with P-gp overexpressed.

^bAntiproliferative activity expressed as IC_{50} values for each cell line, the concentration of compound that caused 50% reduction relative to untreated cells determined by the SRB assay.

^cSI = non-MDR(IC_{50})/MDR(IC_{50}).

^dPaclitaxel.

relative configurations in 3 were determined from NOESY data (Figure 8), showing correlations of H-15/H-6 and H-14, H-6/H-8, CH₃-6'/H-7 and H-5, H-5/H-1, and CH₃-8'/H-2'. The absolute configurations were determined by ECD calculations (Figure 6).

Meroterpenoids make up an exceptional class of structurally diverse natural products formed by the hybridization of terpenoids and nonterpenoids. Set While few meroterpenoids have been isolated from plants, this compound type is found abundantly in bacteria, fungi, algae, and marine organisms. The most common meroterpenoids are hybrids of terpenoids with polyketides and, to a lesser extent, flavonoids. Although small numbers of meroterpenoids containing flavonoid and cyclic terpenoid components have been reported, in all of them, both components were connected simply. In contrast, complex connections were observed between terpenoids and aromatic polyketides, such as phloroglucinol, syncarpic acid, chromane/chromene, and others.

The newly isolated syzygioblanes A–C (1–3, respectively) could presumably be biosynthesized through a [4+2] cyclization between the 6,8-dimethylflavanone desmethoxymatteucinol (4), which was also isolated from *S. oblanceolatum* in this study, and an appropriate germacrane-based bi- or tricyclic sesquiterpene with an exo-olefin (Figure 9). Fungal meroterpenoids, xenovulenes, were also found to be biosynthesized through an intermolecular hetero [4+2] cyclization between humulene, a sesquiterpene, and tropolones. 10

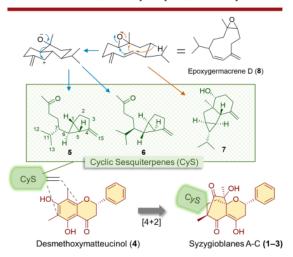


Figure 9. Possible biosynthetic pathway.

Compound 5 and its epimer, 6, with an exo-olefin on a bicyclo[3.1.0]hexane, which might be biosynthesized through epoxygermacrene-D (8), ¹¹ are the necessary sesquiterpene precursors to 1 and 2, respectively, but they have not yet been isolated from nature. Sesquiterpene 7, the needed sesquiterpene component for 3, is a diastereomer of torienol. ¹² Compounds 5–7 were previously synthesized by the treatment of 8 with basic alumina. ^{11,13}

Isolated compounds 1-3 and 4 for comparison were evaluated for antiproliferative activity against five human tumor cell lines (HTCLs): A549 (lung adenocarcinoma), MDA-MB-231 (triple-negative breast cancer), MCF-7 (HER2negative breast cancer), KB (HeLa derivative, originally isolated from epidermoid), and KB-VIN (vincristine resistant KB subline with P-gp overexpressed) (Table 3). Compounds 1 and 2 exhibited potent antiproliferative activity against all tested HTCLs with IC $_{50}$ values of 0.42–13.0 $\mu\mathrm{M}.$ Interestingly, compounds 1-3 showed greater inhibition of the growth of KB-VIN multidrug resistant cells than of the KB chemosensitive parental subline. In particular, compound 1 showed the most hypersensitivity against KB-VIN with a selective index (SI, the IC50 value against KB divided by the IC50 value against KB-VIN) of 14.5. This peculiar phenomenon, greater effectiveness against MDR cells than against drug-sensitive cells, is termed collateral sensitivity (CS). 14 In a recent review, 15 ~70 plant-derived natural products showing CS against drug resistant cancer cells were cited; however, natural products with SI values of >10 and a nanomolar IC50 value against MDR cell lines are rare.

■ ASSOCIATED CONTENT

Data Availability Statement

The data underlying this study are available in the published article and its Supporting Information.

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.orglett.4c01248.

Experimental procedures, spectroscopic data, ORTEP structure for syzygioblane B (2), X-ray structure reports, standard deviation (SD) for antiproliferative activity, and information for the conformers used for ECD calculations (PDF)

Accession Codes

CCDC 2344822 and 2344826 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The

Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

AUTHOR INFORMATION

Corresponding Author

Kyoko Nakagawa-Goto — School of Pharmaceutical Sciences, College of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa 920-1192, Japan; Chemical Biology and Medicinal Chemistry, UNC Eshelman School of Pharmacy, The University of North Carolina, Chapel Hill, North Carolina 27599-7568, United States; orcid.org/0000-0002-1642-6538; Email: kngoto@p.kanazawa-u.ac.jp

Authors

Nona Koga — School of Pharmaceutical Sciences, College of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa 920-1192, Japan

Yohei Saito — School of Pharmaceutical Sciences, College of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa 920-1192, Japan

Katsunori Miyake — School of Pharmacy, Tokyo University of Pharmacy and Life Sciences, Hachioji, Tokyo 192-0392, Japan

Saidanxia Amuti — School of Pharmaceutical Sciences, College of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa 920-1192, Japan

Shuichi Fukuyoshi — School of Pharmaceutical Sciences, College of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa 920-1192, Japan

Satoshi Yoshida — Department of Applied Chemistry, School of Engineer 23, The University of Tokyo, Kashiwa, Chiba 277-0882, Japan

Sota Sato — Department of Applied Chemistry, School of Engineering, T2 University of Tokyo, Kashiwa, Chiba 277-0882, Japan; Institute for Molecular Science, National Institutes 2 Natural Sciences, Okazaki, Aichi 444-8787, Japan; orcid.org/0000-0002-7395-2112

Yusuke Yamada — Structural Biology Research Center, Institute of Materials Structure Science, High Energy Accelerator Research Organization (KEK), Tsukuba, Ibaraki 305-0801, Japan; orcid.org/0000-0002-2651-4005

Akihito Ikeda — Structural Biology Research Center, Institute of Materials Structure Science, High Energy Accelerator Research Organization (KEK), Tsukuba, Ibaraki 305-0801, Japan

Naruhiko Adachi — Structural Biology Research Center, Institute of Materials Structure Science, High Energy Accelerator Research Organization (KEK), Tsukuba, Ibaraki 305-0801, Japan; Life Science Center for Survival Dynamics, Tsukuba Advanced Research Alliance (TARA), University of 2 Tsukuba, Tsukuba, Ibaraki 305-8577, Japan

Masato Kawasaki — Structural Biology Research Center, Institute of Materials Structure Science, High Energy Accelerator Research Organization (KEK), Tsukuba, Ibaraki 305-0801, Japan

Akira Takasu — Structural Biology Research Center, Institute of Materials Structure Science, High Energy Accelerator Research Organization (KEK), Tsukuba, Ibaraki 305-0801, Japan; Life Science Center for Survival Dynamics, Tsukuba Vanced Research Alliance (TARA), University of Tsukuba, Tsukuba, Ibaraki 305-8577, Japan

Shinji Aramaki — Life Science Center for Survival Dynamics, Tsukuba 12 vanced Research Alliance (TARA), University of Tsukuba, Tsukuba, Ibaraki 305-8577, Japan; Tietz Video and Image Processing Systems Japan G.K., Minato-ku, Tokyo 105-1003, Japan

Toshiya Senda – Structural Biology Research Center, Institute of Materials Structure Science, High Energy Accelerator Research Organization (KEK), Tsukuba, Ibaraki 305-0801, Iapan

Abdul Rahim — School of Pharmaceutical Sciences, College of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa 920-1192, Japan; Department of Pharmaceutical Sciences and Technology, Faculty of Pharmacy, Hasanuddin University, Makassar 90245, Indonesia

Ahmad Najib — School of Pharmaceutical Sciences, College of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa 920-1192, Japan; orcid.org/0000-0001-5055-7964

Gemini Alam – Department of Pharmaceutical Sciences and Technology, Faculty of Pharmacy, Hasanuddin University, Makassar 90245, Indonesia

Nobuyuki Tanaka — Department of Botany, National Museum of Nature and Science, Tsukuba, Ibaraki 305-0005, Japan

Complete contact information is available at: https://pubs.acs.org/10.1021/acs.orglett.4c01248

Note

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The authors thank Dr. Wuu-Kuang Soh at the University of Dublin (Dublin, Ireland) for confirmation of the identification. The authors appreciate the critical comments, suggestions, and editing of the manuscript by Dr. Susan L. Morris-Natschke (The University of North Carolina at Chapel Hill). This work was supported by JSPS KAKENHI Grant 16H05811 and the Mitani Foundation for Research and Development (awarded 2 K.N.-G.). MicroED experiments were supported by the Research Support Project for Life Science and Drug Discovery [Basis for Supporting Innovative Drug Discovery and Life Science Research (BINDS)] from AMED under Grant JP22ama121001 to T.S. and Grant JP23ama121005 to S.S. Synchrotron X-ray diffraction experiments at BL-17A at KEK PF ring were supported by the Photon Factory Program Advisory Committee under Grants 2021G589 and 2023G604.

■ REFERENCES

(1) Ashton, P. Notes on northern Borneo Myrtraceae. Sandakania 2016, 22, 55-75.

(2) Low, Y. W.; Rajaraman, S.; Tomlin, C. M.; Ahmad, J. A.; Ardi, W. H.; Armstrong, K.; Athen, P.; Berhaman, A.; Bone, R. E.; Cheek, M.; Cho, N. R. W.; Choo, L. M.; Cowie, I. D.; Crayn, D.; Fleck, S.; Ford, A. J.; Forster, P. I.; Girmansyah, D.; Goyder, D. J.; Gray, B.; Heatubun, C. D.; Ibrahim, A.; Ibrahim, B.; Jayasinghe, H. D.; Kalat, M. A.; Kathriarachchi, H. S.; Kintamani, E.; Koh, S. L.; Lai, J. T. K.; Lee, S. M. L.; Leong, P. K. F.; Lim, W. H.; Lum, S. K. Y.; Mahyuni, R.; McDonald, W. J. F.; Metali, F.; Mustaqim, W. A.; Naiki, A.; Ngo, K. M.; Niissalo, M.; Ranasinghe, S.; Repin, R.; Rustiami, H.; Simbiak, V. I.; Sukri, R. S.; Sunarti, S.; Trethowan, L. A.; Trias-Blasi, A.; Vasconcelos, T. N. C.; Wanma, J. F.; Widodo, P.; Wijesundara, D. S. A.; Worboys, S.; Yap, J. W.; Yong, K. T.; Khew, G. S.; Salojärvi, J.;

- Michael, T. P.; Middleton, D. J.; Burslem, D. F. R. P.; Lindqvist, C.; Lucas, E. J.; Albert, V. A. Genomic insights into rapid speciation within the world's largest tree genus *Syzygium*. *Nat. Commun.* **2022**, *13*, 5031.
- (3) (a) Syzygioblane A (1): colorless amorphous solid; $[\alpha]^{25}_{D}$ –54.8 (c 0.10, CHCl₃); IR (neat) ν_{max} 2925, 2871, 2360, 2342, 1717, 1710, 1663, 1603, 1457, 1367, 1319, 1283, 1160, 1140, 1070, 990, 933, 917, 850, 751, 700, 669 cm⁻¹; ¹H and ¹³C NMR data in Tables 2 and 3; HRFABMS m/z 521.2893 [M + H]⁺ (calcd for $C_{32}H_{41}O_{6t}$ 521.2903). (b) Syzygioblane B (2): colorless amorphous solid; $[\alpha]^{25}_{\rm D}$ –96.2 (c 0.10, CHCl₃); IR (neat) ν_{max} 3404, 2952, 2871, 2360, 2345, 1725, 1696, 1660, 1602, 1456, 1365, 1322, 1276, 1215, 1185, 1146, 1097, 1049, 989, 934, 924, 850, 816, 760, 701, 669, 599, 529, 464 cm⁻¹; ¹H and 13C NMR data in Tables 2 and 3; HRFABMS m/z 521.2871 [M + H]⁺ (calcd for C₃₂H₄₁O₆, 521.2903). (c) Syzygioblane C (3): pale yellow oil; $[\alpha]^{25}_{D}$ -76.7 (c 0.10, CHCl₃); IR (neat) ν_{max} 2951, 2872, 2360, 2342, 1730, 1660, 1603, 1453, 1363, 1309, 1217, 1139, 1077, 1034, 856, 749, 699, 669 cm⁻¹; ¹H and ¹³C NMR data in Tables 2 and 3; HRFABMS m/z 521.2871 [M + H]+ (calcd for C32H41O61 521.2 203).
- (4) Nannenga, B. L.; Gonen, T. The cryo-EM method microcrystal electron diffraction (MicroED). Nat. Methods 2019, 16, 369–379.
- (5) Goyer, E.; Lavaud, C.; Massiot, G. Meroterpenoids? A historical and critical review of this biogenetic determinant. *Nat. Prod. Rep.* 2023, 40, 1071–1077.
- (6) Fuloria, N. K.; Raheja, R. K.; Shah, K. H.; Oza, M. J.; Kulkarni, Y. A.; Subramaniyan, V.; Sekar, M.; Fuloria, S. Biological activities of meroterpenoinds isolated from different sources. Front. Pharm. 2022, 13, 830103.
- (7) Han, J.; Jiang, L.; Zhang, L.; Quinn, R. J.; Liu, X.; Feng, Y. Peculiarities of meroterpenoids and their bioproduction. *Appl. Microbio. Biothech.* **2021**, *105*, 3987–4003.
- (8) Nazir, M.; Saleem, M.; Tousif, M. I.; Anwar, M. A.; Surup, F.; Ali, I.; Wang, D.; Mamadalieva, N. Z.; Alshammari, E.; Ashour, M. L.; Ashour, A. M.; Ahmed, I.; Elizbit; Green, I. R.; Hussain, H. Meroterpenoids: A comprehensive update insight on structural diversity and biology. *Biomolecules* **2021**, *11*, 957.
- (9) (a) Long, C.; Marcourt, L.; Raux, R.; David, B.; Gau, C.; Menendez, C.; Gao, M.; Laroche, M.-F.; Schambel, P.; Delaude, C.; Ausseil, F.; Lavaud, C.; Massiot, G. Meroterpenes from Dichrostachys cinerea Inhibit Protein Farnesyl Transferase Activity. J. Nat. Prod. 2009, 72, 1804-1815. (b) Kenmogne, C. F.; Eckhardt, P.; Tchuenguem, R. T.; Groβ, J.; Ngouafong, F.; Ponou, B. K.; Dzoyem, J. P.; Teponno, R. B.; Opatz, T.; Tapondjou, L. A. Diplomeroterpenoid G: An unusual meroterpenoid from Mimosa pudica Linn. (Mimosaceae). Tetrahedron Lett. 2023, 120, 154451. (c) Zhu, Y. D.; Wu, H.-F.; Ma, G.-X.; Chen, R.-C.; Long, H.-L.; Zuo, Z.-L.; Luo, Y.; Zhu, N.-L.; Hou, B.; Xu, X.-D.; Sun, G.-B.; Yang, J.-S. Clinoposides A-F: meroterpenoids with protective effects on H9c2 cardiomyocyte from Clinopodium chinense. RSC Adv. 2016, 6, 7260-7266. (d) Zhu, Y. D.; Chen, R.-C.; Wang, H.; Jiang, H.; Huang, X.-L.; Zhang, M.-L.; Li, L.-Y.; Hu, Z.; Xu, X.-D.; Wang, C.-J.; Ye, X.-X.; Yang, J.-S. Two new flavonoid-triterpene saponin meroterpenoids from Clinopodium chinense and their protective effects against anoxia/ reoxygenation-induced apoptosis in H9c2 cells. Fitoterapia 2018, 128, 180 - 186.
- (10) (a) Schor, R.; Schotte, C.; Wibberg, D.; Kalinowski, J.; Cox, R. J. Nat. Commun. 2018, 9, 1963. (b) Schotte, C.; Li, L.; Wibberg, D.; Kalinowski, J.; Cox, R. J. Angew. Chem. Int. 2020, 59, 23870–23878. (11) Niwa, M.; Iguchi, M.; Yamamura, S. Biomimetic reactions of germacrenes. Tetrahedron Lett. 1979, 20, 4291–4294.
- (12) Itokawa, H.; Matsumoto, H.; Mihashi, S. Isolation of oppositane- and cyaloeudesmane-type sesquiterpenoids from *Torilis japonica* D. C. Chem. Lett. 1983, 12, 1253–1256.
- (13) Yamamura, S.; Niwa, M.; Ito, M.; Saito, Y. Biomimetic conversion of epoxygermacrene-D to a novel sesquiterpene of the plant *Torilis japonica DC. Chem. Lett.* 1982, 11, 1681–1684.

- (14) Pluchino, K. M.; Hall, M. D.; Goldsborough, A. S.; Callaghan, R.; Gottesman, M. M. Collateral sensitivity as a strategy against cancer multidrug resistance. *Drug Resist. Updat.* **2012**, *15*, 98–105.
- (15) Efferth, T.; Saeed, M. E. M.; Kadioglu, O.; Seo, E.-J.; Shirooie, S.; Mbaveng, A. T.; Nabavi, S. M.; Kuete, V. Collateral sensitivity of natural products in drug-resistant cancer cells. *Biotechnol. Adv.* 2020, 38, 107342.

Cyclic Sesquiterpene–Flavanone [4+2] Hybrids, Syzygioblanes A–C, Found in an Indonesian Traditional Medicine, "Jampu Salo" (Syzygium oblanceolatum)

ORIGINALITY REPORT

9% SIMILARITY INDEX

5%
INTERNET SOURCES

9%
PUBLICATIONS

5% STUDENT PAPERS

< 5%

PRIMARY SOURCES

pesquisa.bvsalud.org

Publication

5%

Eisuke Tsunekawa, Yusuke Otsubo, Yusuke Yamada, Akihito Ikeda et al. "X-ray and Electron Diffraction Observations of Steric Zipper Interactions in Metal-Induced Peptide Cross-β Nanostructures", Journal of the American Chemical Society, 2023

Exclude quotes On Exclude matches

Exclude bibliography On